

# PRELIMINARY REPORT ON NINETEEN DIGESTION METHODS TESTED ON VARIOUS GEOCHEMICAL EXPLORATION SAMPLE MEDIAS

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PRELIMINARY REPORT ON NINETEEN DIGESTION METHODS TESTED ON VARIOUS GEOCHEMICAL EXPLORATION SAMPLE MEDIAS

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## I. INTRODUCTION

The extractability of metal ions and selectivity of certain sample components by various digestion methods is an important consideration in exploration geochemistry. This subject has been discussed by many authors, some of which are given in the reference section at the back of this report.

For this study, nineteen different digestion methods were tested on seven different types of geochemical samples and analyzed for copper, nickel and zinc. The digestion methods are described in the Appendix. The solutions were analyzed by A. W. Klaysmat (Minerals Division Chemist) on a Perkin-Elmer 303 atomic absorption spectrophotometer. The results of these tests are considered preliminary. The results for each digestion method and sample media are presented in Tables 1, 2 and 3.

# II. SOME OBSERVATIONS - MINERAL ACID DIGESTION METHODS

In order to compare the extractability of various mineral acid digestion methods, the concentrations of each metal for each sample type and for each digestion method was divided by the concentrations obtained for the (HF, HNO<sub>3</sub> & HCl) digestion. This digestion is not total, but is nearly total. The results of the "percent of total metal" are presented in Table 4.

The <u>hot</u> methods usually extracted substantially more metal than the <u>cold</u> methods, as would be expected. However, for the Gyttja sample, the difference between hot and cold methods is small. The <u>aqua regia</u> methods and <u>HC1</u> methods extract less metal from the <u>silicates</u> than the <u>HNO<sub>3</sub></u> methods as evidenced by the Ni for the Troctolite.  $HNO_3$  will extract a significant portion of metals in silicates (Timperley and Allan, 1974). HCl does not attack silicates to the extent of  $HNO_3$  (Timperley and Allan, 1974). HCl generally attacks only the less resistant silicates (Bradshaw et al., 1974).

<u>Aqua regia</u> and <u>HNO3</u> generally extract more metal from the <u>sulfides</u> than <u>HC1</u> as evidenced by Cu for the Troctolite (Mineralized) and Troctolite.

For the <u>organic</u> samples, A-horizon and Gyttja, the various <u>HNO<sub>3</sub></u> methods extract slightly more metal than the various <u>aqua regia</u> methods. The <u>HC1</u> and HNO<sub>3</sub> methods generally extract about the same amount of metal from the organics. HCl will extract significant amounts of metals from organics (Timperley and Allan, 1974). HNO<sub>3</sub> is known to decompose organic matter (Timperley and Allan, 1974; Rose, 1975). The HF, HNO<sub>3</sub>, and HCl digestion, except for some Cu values extracted more metal than the other methods in the Gyttja sample, which suggests silicates are present in the gyttja.

For the samples containing <u>Fe-Mn hydroxides</u>, B-horizon and Fe-Mn Hydroxide, <u>HC1</u> methods tend to extract slightly more metal than the <u>HNO<sub>3</sub></u> methods. HC1 is generally known to extract more metal from Fe-Mn hydroxides than  $HNO_3$  (Rose, 1975). The C-horizon sample contains lesser amounts of Fe-Mn hydroxides, but can be considered in this comparison. The HC1 methods extracted more metal than the aqua regia methods.

### III. SOME OBSERVATIONS - WEAK EXTRACTION METHODS

The results of the weak extraction methods expressed as "percent of total metal" is presented in Table 5.

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A. 0.5M HC1

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The <u>hot</u> method does extract significantly more metal than the cold method.

The 0.5M HCl leach does appear to extract a relatively large amount of Fe-Mn hydroxide and <u>organic</u> held metal as evidenced by the Fe-Mn Hydroxide and Gyttja samples respectively.

It does appear to extract significant amounts of metal held in <u>clay minerals</u> as shown for the C-horizon sample, but a large portion of this metal may be held in Fe-Mn hydroxides. This method, also, did not extract significant amounts of <u>sulfide</u> held Cu from the mineralized Troctolite sample as compared to the other samples. EDTA

The EDTA method did extract a large portion of the metal in the organics: A-horizon and Gyttja samples.

It did not extract any of the <u>sulfide</u> held Cu in the mineralized Troctolite sample.

Only a small portion of the <u>Fe-Mn hydroxide</u> held metal in the B-horizon sample was extracted. However, a large portion of metal was extracted in the Fe-Mn hydroxide sample.

For the C-horizon sample, the EDTA method did extract a large portion of probable clay held metal or loosely bonded metal in the Fe-Mn hydroxides. EDTA will complex exchangeable ions in clay minerals (Rose, 1975).

EDTA, a chelating extractant, does not remove significant metal from silicates. EDTA is mainly specific for organic held metal (Timperley and Allan, 1974).

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#### C. Ammonium Citrate/Hydrogen Peroxide

Ammonium citrate reduces iron hydroxides.

<u>Hydrogen peroxide</u> is known to dissolve <u>Mn Hydroxides</u> (Rose, 1975). Also, hydrogen peroxide is a strong oxidizing agent and, therefore, is known to dissolve organics and sulfides (Rose, 1975).

The observed extraction of metals in the samples containing <u>Fe-Mn hydroxides</u>, B-horizon and Fe-Mn hydroxide samples, probably results from the reduction of iron by the ammonium citrate and dissolution of Mn hydroxides by the hydrogen peroxide. The extracted metal in the C-horizon sample may be the result of hydrogen peroxide acting on the Mn hydroxides and ammonium citrate on the Fe hydroxides.

A significant portion of metal was extracted from the <u>organics</u> in the Gyttja sample. However, a portion of this metal is probably contained in Fe-Mn hydroxides, extractable by the ammonium citrate and hydrogen peroxide and in sulfide form, extractable by the hydrogen peroxide. The A-horizon sample did not indicate that significant organic held metal was extracted by this method.

The Troctolite (mineralized) and Troctolite samples indicate that large portions of <u>sulfide</u> held metal is extracted by the hydrogen peroxide.

## D. Hydroxylamine Hydrochloride/Ammonium Citrate

Ammonium citrate reduces <u>iron hydroxides</u> and hydroxylamine hydrochloride Mn hydroxides (Rose, 1975).

This method appears to extract minor amounts of <u>organic</u> and sulfide held metal as evidenced by the A-horizon, Gyttja, Troctolite (mineralized) and Troctolite samples. The higher percents of extracted Ni and Zn for the Gyttja sample probably are from Fe-Mn hydroxides.

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The B-horizon, C-horizon and Fe-Mn hydroxide samples, as compared to the other samples, indicate that this method is essentially specific for <u>Fe-Mn hydroxides</u>.

## E. Ascorbic Acid/Hydrogen Peroxide

This method is essentially specific for <u>sulfide</u> held metal as compared to <u>hydroxide</u> held metal (Gunton and Nichol, 1974). However, the hydrogen peroxide will attack the organics.

The extraction of organic held metal by the hydrogen peroxide is illustrated by the A-horizon, B-horizon and Gyttja samples. The B-horizon sample generally will contain organic material washed down from the A-horizon.

The almost complete lack of extraction of <u>Fe-Mn hydroxide</u> held metal is evidenced by the C-horizon and Fe-Mn hydroxide samples.

A fair amount of <u>sulfide</u> held metal was extracted from the Troctolite (mineralized) and Troctolite samples, but it is less than expected. A portion of the metal in the Gyttja sample is probably extracted from sulfide in addition to the organic held metal.

F. Evaluation and Comparison of Weak Extraction Methods

Based on the above mentioned results and literature survey, Table 6 has been prepared on the apparent relative extractability of weak extraction methods.

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	<u>A Horizon</u>	B Horizon (Rainy Lobe)	C Horizon (Indus Till)	Fe-Mn Hydroxide	Gyttja	*Troctolite (Mineralized)	*Troctolite
AN-1(HF, NHO <sub>3</sub> & HC1)	28	71	20	122	28	5265	95
HNO <sub>3</sub> & HC1 Cold	8	35	16		27	5180	85
HNO <sub>3</sub> & HCl Hot	20	56	18		28	4880	86
HNO3 Cold	9	21	12°		27	1920	60
HNO <sub>3</sub> Hot	10	50	19		28	5600	92
HCl Cold	10	45	18		25	1260	58
HCl Hot	13	68	22		25	4850	92
4M $HNO_3$ & 1M HC1 Cold	8	21	13	64	17	1820	44
4M HNO <sub>3</sub> & 1M HC1 Hot	12	54	20	119.	23	4820	86
4M HNO <sub>3</sub> Cold	10	24	12	56	18	250	35
4M HNO <sub>3</sub> Hot	9	61	19	127	26	4780	84
4M HC1 Cold	8	23	11	72	15	1800	30
4M HC1 Hot	8	55	17	57	16	2500	68
0.5M HC1 Cold	2	6	1	34	4	15	8
0.5M HC1 Hot(AN-12)	2	18	0	43	4	65	18
EDTA	4	5	4	46	12	17	
AMM. Citrate	1	3	2	22	6	1012	41
H.H. Citrate	0.2	5.4	2.8	6.8	0.4	0.4	0.8

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# TABLE 1: COPPER (PPM)

\* -100 mesh fraction; all other samples -80 mesh fraction

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AN-8 Ascorbic Acid/

Hydrogen Peroxide

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TABLE 2: NICKEL (PPM)

	A Horizon	B Horizon (Rainy Lobe)	C Horizon (Indus Till)	Fe-Mn Hydroxide	Gyttja	*Troctolite (Mineralized)	*Troctolite
AN-1(HF, NHO <sub>3</sub> & HC1)	0	96	23	29	48	1270	1020
HNO <sub>3</sub> & HC1 Cold	0	0	0		9	3140	354
HNO <sub>3</sub> & HC1 Hot	0	0	0		0	1990	646
HNO3 Cold	0	0	0		0	990	200
HNO <sub>3</sub> Hot	0	11	0		33	2970	880
HC1 Cold	0	22	11		28	200	334
HCl Hot	0	80	17		45	300	560
4M HNO <sub>3</sub> & 1M HCl Cold	0	0	0	0	0	780	254
4M $HNO_3$ & 1M HCl Hot	0	24	0	0	5	980	601
4M HNO <sub>3</sub> Cold	0	0	0	0	0	178	498
4M HNO <sub>3</sub> Hot	0	54	0	0	30	200	846
4M HC1 Cold	0	0	0	0	0	215	320
4M HC1 Hot	0	45	8	4	25	340	512
0.5M HC1 Cold	4	2	0	6	11	57	38
0.5M HC1 Hot(AN-12)	2	12	4	7	21	96	79
EDTA	0	0	4	6	12	12	
AMM. Citrate	0	0	0	0	0	315	0
H.H. Citrate	1.6	0.2	6.6	2.8	5	27	12
AN-8 Ascorbic Acid/ Hydrogen Peroxide	0	0	0	0	0	9	42

 $\star$  -100 mesh fraction; all other samples -80 mesh fraction

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# TABLE 3: ZINC (PPM)

	<u>A Horizon</u>	B Horizon (Rainy Lobe)	C Horizon (Indus Till)	Fe-Mn Hydroxide	Gyttja	*Troctolite (Mineralized)	*Troctolite_
AN-1(HF, NHO <sub>3</sub> & HCl)	66	76	48	88	101	210	96
HNO3 & HCl Cold	10	16	19		83	57	41
HNO <sub>3</sub> & HCl Hot	14	36	26		94	70	62
HNO <sub>3</sub> Cold	30	1	4		68	41	10
HNO <sub>3</sub> Hot	59	25	22		99	98	60
HC1 Cold	6	25	22		83	68	45
HC1 Hot	71	47	35		98	110	75
4M HNO <sub>3</sub> & 1M HCl Cold	6	. 8	15	42	68	67	30
4M $HNO_3$ & 1M HCl Hot	16	34	27	58	89	104	۲0 ا ∞
4M HNO <sub>3</sub> Cold	8	10	20	40	64	61	38 1
4M HNO <sub>3</sub> Hot	19	41	33	57	98	105	66
4M HC1 Cold	10	13	15	42	81	74	. 46
4M HCl Hot	19	41	35	57	89	104	73
0.5M HC1 Cold	7	1	0	20	52	8	5
0.5M HC1 Hot(AN-12)	4	6	0	26	65	17	13
EDTA	23	0	3	112	176	40	
AMM. Citrate	2	4	1	14	44	350	2
H.H. Citrate	0.6	. 0	0	3.0	11	0	0
AN-8 Ascorbic Acid/ Hydrogen Peroxide	6	5	0	1	48	480	2

 $\star$  -100 mesh fraction; all other samples -80 mesh fraction

# TABLE 4: PERCENT OF TOTAL METAL FOR EACH MINERAL ACID DIGESTION METHOD (TOTAL METAL = HF, HNO<sub>3</sub> & HC1 DIGESTION)

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	АНо	oriz	on		oriz ny L	on obe)		oriz us I	on 111)	Hy	Fe-M drox		G	yttj	а		octol neral	ite ized)	Tro	octo1	ite
			Zn		<u>Ni</u>			<u>Ni</u>	Zn	•		Zn		Ni			<u>Ni</u>		Cu	<u>Ni</u>	Zn
HNO <sub>3</sub> & HC1 Hot	71	*	21	79	0	47	90	0	54				100	0	93	93	157	33	91	63	65
HNO <sub>3</sub> Hot	36	*	89	70	11	33	95	0	46				100	69	98	106	234	47	97	86	63
HCl Hot	46	*	108	96	83	62	110	74	73				89	94	97	92	24	52	97	55	78
4M HNO <sub>3</sub> & 1M HCl Hot	43	*	24	76	25	45	100	0	56	98	0	66	82	10	88	92	77	50	91	59	73
4M HNO <sub>3</sub> Hot	32	*	29	<sup>.</sup> 86	56	54	95	0	69	104	0	65	93	63	97	91	16	50	88	83	69
4M HCl Hot	29	*	29	77	47	54	85	35	73	47	14	65	57	52	88	47	27	50	72	50	76
0.5M HC1 Hot(AN-12)	7	*	6	25	13	8	0	17	0	35	24	30	14	44	64	1.2	8	8	19	8	14 I 9
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HNO <sub>3</sub> & HC1 Cold	29	*	15	49	0	21	80	0	40				96	19	82	98	247	27	89	35	43
HNO3 Cold	32	*	45	30	0	1	60	0	8				96	0	67	36	78	20	63	20	10
HCl Cold	36	*	9	63	23	33	90	48	46				89	58	82	24	16	32	61	33	47
4M HNO <sub>3</sub> & 1M HC1 Cold	29	*	9	30	0	11	65	0	31	52	0	48	61	0	67	35	61	32	46	25	31
4M HNO3 Cold	36	*	12	34	0	13	60	0	42	46	0	45	64	0	40	5	14	29	37	49	40
4M HCl Cold	29	*	15	32	0	17	55	0	31	59	0	48	54	0	80	34	17	35	32	31	48
0.5M HC1 Cold	7	*	11	8	2	1	5	0	0	28	21	23	14	23	51	.3	4	4	8	4	5

 $\star$  Ni for A-Horizon sample was zero except for 0.5M HCl extractions

# TABLE 5: PERCENT OF TOTAL METAL FOR EACH WEAK EXTRACTION METHOD (TOTAL METAL = HF, $HNO_3$ & HC1 EXTRACTION)

				ВН	oriz	on	СН	loriz	on		Fe-M	'n				Tr	octo	lite				
	A Ho	oriz	on	(Rai	ny L	obe)	(Ind	us T	'ill)	Hy	drox	ide	G	yttj	а	(Mir	leral	ized)	Tro	ctol	ite	
	Cu	<u>Ni</u>	Zn	Cu	Ni	Zn	Cu	Ni	Zn	Cu	<u>Ni</u>	Zn	Cu	Ni	Zn	Cu	<u>Ni</u>	Zn	Cu	<u>Ni</u>	Zn	
0.5M HC1 Cold	7	*	11	. 8	2	1	5	0	0	28	21	23	14	23	51	.3	4	4	8	4	5	
0.5M HC1 Hot(AN-12)	7	*	6	25	13	8	0	17	0	35	24	30	14	44	64	1.2	8	8	19	8	14	
EDTA	14	*	35	7'	0	0	20	17	6	38	21	127	43	25	174	0	1	19				
Amm. Citrate	4	×	3	4	0	5	10	0	2	18	0	16	21	0	44	19	25	167	43	0	2	
H.H. Citrate	1	*	1	8	0	0	14	29	0	6	10	3	1	10	11	0	2	0	1	1	0	
AN-8 Ascorbic Acid/ Hydrogen Peroxide	7	*	9	10	0	7	0	0	0	1	0	1	14	0	48	6	1	229	21	4	2	- 10 -

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\*Zero concentration for total metal

	Organics	Fe-Mn Hydroxides	<u>Clay Minerals</u>	Sulfides	Silicates
0.5M HC1	V	V	G	L	L
EDTA	V	G	G	L	L
Amm. Citrate/ HP	V	V	G	V	F
Amm. Citrate/ HH	L	V	F	L	L
Ascorbic Acid/ HP	v	L	G	V	F

# TABLE 6: RELATIVE EXTRACTIBILITY OF WEAK EXTRACTION METHODS FOR VARIOUS SAMPLE TYPES

Extractibility:	V	-	very	good
	G	-	good	
	$\mathbf{F}$	-	fair	
	L	-	low	

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## APPENDIX: DIGESTION PROCEDURES

- 1. HF,  $HNO_3$  and HC1 (concentrated) Hot (90°C)
- 2.  $HNO_3$  and HC1 Cold

1.0000gm sample digested with 10mls of concentrated HNO<sub>3</sub> and 10mls of concentrated HC1. Final volume in the analysis was 100mls. The sample was kept at room temperature for two hours and then filtered through 40 Whatman filter paper.

3. HNO<sub>3</sub> and HC1 - Hot

1.0000gm sample digested with 10mls of concentrated HNO3 and 10mls of concentrated HC1. Final volume in the analysis was 100mls. The sample was digested at 90°C for two hours and then filtered through 40 Whatman filter paper.

4.  $HNO_3 - Cold$ 

1.0000gm sample was digested with 20mls of concentrated  $HNO_3$ . The final volume in the analysis was 100mls. The sample was digested at room temperature for two hours and then filtered through 40 Whatman filter paper.

5.  $HNO_3 - Hot$ 

1.0000gm sample was digested with 20mls of concentrated  $HNO_3$ . The final volume in the analysis was 100mls. The sample was digested at 90°C for two hours and then filtered through 40 Whatman filter paper.

6. HCl - Cold

1.0000gm sample digested with 20mls of concentrated HC1. The final volume in the analysis was 100mls. The sample was digested at room temperature for two hours and then filtered through 40 Whatman filter paper.

#### 7. HC1 - Hot

1.0000gm sample digested with 20mls of concentrated HC1. The final volume in the analysis was 100mls. The sample was digested at 90°C for two hours and then filtered through 40 Whatman filter paper.

8. 4M HNO3 and 1M HC1 - Cold

1.0000gm sample digested with 10mls of 4M HNO<sub>3</sub> and 10mls of 1M HC1. The final volume in the analysis was 100mls. The sample was digested at room temperature for two hours and then filtered through 40 Whatman filter paper.

9. 4M HNO3 and 1M HC1 - Hot

1.0000gm sample digested with 10mls of 4M  $\rm HNO_3$  and 10mls of 1M HC1. The final volume in the analysis was 100mls. The sample was digested at 90°C for two hours and then filtered through 40 Whatman filter paper.

10. 4M  $HNO_3$  - Cold

1.0000gm sample digested with 20mls of 4M  $HNO_3$ . The final volume in the analysis was 100mls. The samples were digested at room temperature for two hours and then filtered through 40 Whatman filter paper.

11. 4M  $HNO_3$  - Hot

1.0000gm sample digested with 20mls of 4M  $\rm HNO_3$ . The final volume in the analysis was 100mls. The sample was digested at 90°C for two hours and then filtered through 40 Whatman filter paper.

12. 4M HC1 - Cold

1.0000gm sample digested with 20mls of 4M HCl. The final volume in the analysis was 100mls. The samples were digested at room temperature for two hours and then filtered through 40 Whatman filter paper.

# 13. 4M HC1 - Hot

1.0000gm sample digested with 20mls of 4M HC1. The final volume in the analysis was 100mls. The sample was digested at  $90^{\circ}$ C for two hours and then filtered through 40 Whatman filter paper.

14. 0.5M HC1 - Cold

3.000gms sample digested with 20mls of 0.5M HC1. The final volume in the analysis was 100mls. The samples were digested at room temperature for two hours and then filtered through 40 Whatman filter paper.

15. 0.5M HC1 - Hot

3.0000gms sample

16. EDTA

Dissolved 37.22gms EDTA disodium salt in 500mls of distilled water using a 1000ml beaker. The pH was 4.3 and it was adjusted to 4.8 using ammonium hydroxide. Dilute this solution to one liter using 1000ml volumetric flask.

Digestion procedure: 1.0000gm sample was digested for 18 hours stirring every half hour. 15ml of EDTA solution was added to the 1gm sample. After 18 hours, the sample was diluted to 100mls with deionized water and filtered through 40 Whatman filter paper.

17. Citrate

Dissolve 50.0gm ammonium citrate and 50.0gms hydroxylamine hydrochloride in 300mls distilled water using a 1000ml beaker. The pH was 3.7 adjusted to 4.3 using ammonium hydroxide. Dilute this solution to 500mls in a volumetric flask.

Digestion procedure: 5.0000gm sample was digested for 18 hours stirring every half hour. 50.0mls of ammonium citrate solution was added to the 5.0gm sample. After 18 hours, the sample was diluted to 100mls with deionized water and filtered through 40 Whatman filter paper.

- Ascorbic Acid/Hydrogen Peroxide
   Used 5.000gm sample.
- 19. Ammonium Citrate Digestion

Weigh .5000gm sample. Add to 40mls of a 10% w/w ammonium citrate and 20mls hydrogen peroxide 30% (conc.). Shake or stir every half hour for 18 hours. Filter through 40 Whatman filter paper into a flask. Wash carefully with distilled water. Add 5mls of concentrated HC1 - 12M. Boil for one-half hour or until hydrogen peroxide is gone (slight light color change). Do not take to dryness. Bring to 100ml volume with deionized water in volumetric flask.

#### Comments:

All standards were made up with the same concentration of acids that were used in the digestion procedure. It was very difficult to wet some of the organic samples in the cold digestions. Hence some of the fine organic particles floated on the surface tension of the solutions. When it states in the procedure that the samples were stirred every half hour, no stirring took place during non-working hours in the lab. Some of the samples filtered very slowly (4 hours or more); consequently, sometimes the samples were in contact with the digesting solutions longer than the two hour limit. The mineralized troctolite sample was usually off scale when a 1.0000gm sample was diluted to 100mls; therefore, the concentration had to be diluted before the analysis could be completed. (The original 100ml sample was diluted 5 to 20 times depending on the concentration of the elements of interest). The original standards were still used when this happened; consequently, the standards had a higher concentration of acids than some of the diluted samples. All samples were brought to final volume with deionized water.

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